



HUMAN PREGNANE X RECEPTOR (PXR, NR1I2) ACTIVATION ASSAY SYSTEM

Technical Manual for 384-Well Format Assay
Catalog# DPX2-384-001 and DPX2-384-002

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HUMAN PXR NUCLEAR RECEPTOR ACTIVATION ASSAY, WITH ASSESSMENT OF CELL VIABILITY AND CYP3A SUBSTRATE METABOLISM IN A 384-WELL FORMAT

This technical manual is available in its most current version at: www.puracyp.com. Please contact Puracyp if you have questions concerning the use of the assay system. 760-929-9508 ext. 103 or 113; Email: support@puracyp.com

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I. DESCRIPTION

Puracyp's nuclear receptor activation kits are all-inclusive cell-based assay systems. The kits incorporate mycoplasma-free, stably-transfected cells derived from hepatoma cell lines that have been treated with a cell-arrest agent to prevent further proliferation. Exposure to the cell-arrest agent ensures that the cells in the kits can only be used for a single 384-well assay. The kits also include a cell culture-ready assay plate, optimized cell culture medium, cell dosing medium, a positive control specific for the nuclear receptor, CellTiter-Fluor™ for determining cell viability, luciferase detection reagents and, if so ordered, a P450-Glo™ substrate for assessing induction of CYP450 enzyme activity. The primary application of these assay kits is to identify agents with the ability to activate PXR and/or AhR, thereby inducing drug-metabolizing enzymes and ABC transporters.

Puracyp's *Human PXR Activation Assay System* utilizes DPX2™ cells constructed with a proprietary process for producing stably-transfected cell lines. DPX2™ cells harbor the human PXR gene (NR1I2) and a luciferase reporter gene linked to two promoters identified in the human CYP3A4 gene, namely XREM and PXRE. The DPX2™ cells included in the assay kit remain highly viable upon thawing, and can be immediately dispensed into 384-well plates. There is no need for intermediate spin-and-wash steps. However, determining viability and cell titer is recommended. A list of references describing the use of Puracyp's DPX2™ cells and other stable cell lines is given in this manual.

Puracyp's *Human PXR Activation Assay System* also includes CellTiter-Fluor™, a cell viability detection reagent, and ONE-Glo™, a luciferase detection reagent, from Promega Corporation. Since the luciferase reporter gene is stably integrated into Puracyp's DPX2™ cells, the assay gives extremely low background, high sensitivity, and a broad linear dynamic range typical of bio-luminescence reporter gene technology. Further details on the use of CellTiter-Fluor™, ONE-Glo™ and the P450-Glo™ CYP3A4 Assay with Luciferin-IPA can be found at www.promega.com.

In a typical assay, DPX2™ cells are first dispensed into the wells of the assay plate, and then placed in a CO₂ incubator at 37°C to equilibrate overnight. The following day, the cells are treated with user's test compounds and the appropriate positive control. Since the DPX2™ cells included with the kit are non-proliferative, test treatment should take place within 24 h of plate seeding. Once treated, the cells are returned to the CO₂ incubator at 37°C for 48 h. The dosing medium is then discarded, cell viability is assessed fluorimetrically with CellTiter-Fluor™, and luciferase activity measured using ONE-Glo™. The luminescence light intensity is directly proportional to the extent of PXR activation and accompanying gene transcription in the DPX2™ cells.

II. ASSAY KIT COMPONENTS & STORAGE CONDITIONS

Puracyp's *Human PXR Activation Assay System* (Product DPX2™-384-001 and DPX2™-384-002) contains materials to perform reporter assays in a 384 well plate format. The kits are constructed so that 128 reporter assays can be performed in triplicate. Individual aliquots of DPX2™ cells, media and detection reagents are intended for a single-use. Once DPX2™ cells have been thawed, they can neither be refrozen nor cultured for future use due to the utilization of a cell-arrest agent during cellular preparation. Therefore, if an entire 384-well plate is not seeded, the remaining cells should be discarded

Assay kits are shipped on dry ice and DPX2 cells are shipped in a liquid nitrogen dry shipper. **If the DPX2™ cells will not be used immediately, they require storage in liquid nitrogen.** The other kit components can be stored at -20° C for up to 90 days. Individual expiration dates are provided with each kit.

KIT COMPONENT	AMOUNT	STORAGE TEMPERATURE
Tube A - DPX2™ Cells	6 million	Liquid Nitrogen
Tube B - Positive Control (20 mM)	1 x 10 µl	-20° C
Tube C - Culture Media	1 x 10 mL	-20° C
Tube D - Dosing Media	1 x 35 mL	-20° C
Tube E	1 x 10 mL	-20° C
Tube F - ONE-Glo™ Assay Buffer	1 x 10 mL	-20° C
Tube G - ONE-Glo™ Assay Substrate, Lyophilized	1 vial	-20° C
Tube H - P450-Glo™ Buffer	1x 10 ml	-20° C
Tube I - Luciferin Detection Reagent, Lyophilized	1 vial	-20° C
Tube J - CellTiter-Fluor™	1 x 5 µl	-20° C
Tube K - Luciferin-IPA	1 vial	-20° C
384-Well Microtiter Plate	1	Ambient

III. MATERIALS TO BE SUPPLIED BY USER

The following materials are required by the end-user, and should be available prior to starting the assay procedure.

FOR RECEIPT OF CELLS

- Cell culture-rated hood
- Humidified 5% CO₂/37°C incubator suitable for mammalian cell culture
- 37° C water bath
- Sterile multi-channel media basins or reservoirs
- 70% ethanol mixture
- Multi-channel pipette or liquid handler & sterile tips

FOR DAY 1

- Cell culture-rated hood
- Humidified 5% CO₂/37°C incubator
- 37°C water bath
- Sterile 1.5 ml microfuge tubes for making DMSO dilutions
- Sterile deep 384 well plate (100 µL per well capacity) to make test compound doses (compound dilution plate).
- Media trough or basin
- DMSO or other solvent for use as a test compound diluent and as the negative control
- Multi-channel pipette or liquid handler & sterile tips

FOR DAY 3 (PXR Activation)

- Multi-mode microplate reader with capacity to measure both luminescence and fluorescence.
- 2 x Media troughs
- 15 ml sterile conical centrifuge tube
- Multi-channel pipette or liquid handler & sterile tips

FOR DAY 3 (PXR activation and P450 metabolism)

- **White** 384-well plate (non-sterile) to read luminescence
- Multi-mode microplate reader with capacity to read luminescence and fluorescence.
- 3 x Media troughs
- Multi-channel pipette or liquid handler & sterile tips
- 2 x 15 ml sterile conical centrifuge tube

IV. ASSAY PROCEDURE

NOTE: It is advisable to review the following section before proceeding with the assay

Puracyp's *Human PXR Activation Assay* incorporates one overnight and one 48 h incubation. *Steps 1-11* are performed upon receipt of cells, and require about **30 min** to complete. *Steps 12-21* are performed on Day 1, and need **1.5 - 4.5 h** to complete, depending on the number of dilutions and compounds to be tested. *Steps 22-32* are performed on Day 3, and require **1.5- 3 h** to perform.

RECEIPT OF CELLS: ALL STEPS MUST BE PERFORMED USING PROPER ASEPTIC TECHNIQUE

1. Wipe down tissue culture hood with 70% alcohol solution.
2. Remove 384-well plate from kit and place inside tissue culture hood.
3. Place sterile media trough or basin in hood.
4. Prepare multi-channel pipette or liquid handler with sterile tips.
5. Place Tube C from kit directly into a 37°C water bath until medium has thawed and warmed to 37°C, and transfer 10 ml into a sterile trough.
6. Place Tube A containing the DPX2™ cells into a 37°C water bath for 1-2 min, or until cell suspension has thawed. **Do Not Overheat**
7. Sterilize outside of Tube A containing the cells with 70% alcohol solution. Using a 2 ml pipette, aspirate the cell mixture up and down twice to mix, then transfer cells to the sterile trough containing 10 ml of medium from Tube C.
8. Remove 2 ml of the cell-medium mixture, and use it to rinse out Tube A. Transfer contents back to the media trough.
9. Uniformly mix cells with medium by pipetting up and down at least 10X with a 10 ml pipette. It is recommended that the cells be counted to ensure there are 6 million viable cells/vial.
10. Using the multi-channel pipette or liquid handler, transfer 25 µl of cell media suspension from the trough to the individual wells of 384-well plate. Ensure that the cells remain suspended in medium by agitating the trough between the seeding of each row/column.
11. Place seeded plate into humidified 5% CO₂/37°C incubator overnight.

DAY 1

12. Place Tube B from kit at room temperature.
13. Place Tube D from kit at room temperature, and allow medium to thaw.
14. Wipe down tissue culture hood with 70% alcohol solution.

THE FOLLOWING STEPS ARE PERFORMED IN A STERILE HOOD

15. Prepare positive control (Tube B) dilutions as described in **Appendix 1**.
16. Prepare Test Compound dilutions as described in **Appendix 2** if triplicate determinations are planned. Prepare the vehicle control (1% DMSO) by adding 1 µl of DMSO to 100 µl of media from Tube D. If performing metabolism assay, return remaining contents of Tube D to 4°C. Ten mls of Tube D will be needed for the metabolism assay (see step #24 below).
17. Remove 384-well plate containing the cells from the incubator and place into the hood.
18. Prepare the plate for test agent addition by demarcating on the lid those wells that will receive test articles, positive controls, vehicle control, and medium control.
19. Remove the liquid from each well by gently tapping the plate several times upside down on a sterile surface. Ensure that all media is removed but do not disturb the cells.
20. Gently add 25 µl of medium containing test agent, positive control or vehicle control to the plate wells. Media addition is performed by gently pipetting the liquid against the well walls rather than directly onto the cells attached to the well bottom.
21. Return the plate to the incubator, and continue to incubate at 37°C for 48 h.

DAY 3 PXR Activation (DPX2-384-001 kit)

22. Allow Tubes E, F, G & J to reach room temperature.
23. Add the contents of Tube F (ONE-Glo™ Assay Buffer) to Tube G (ONE-Glo™ Assay Substrate), cap, and invert the bottle 3X to mix.
24. Transfer 10 ml from Tube E to a sterile 15 ml conical tube, and add 5 µl from Tube J (CellTiter-Fluor™). Mix by inverting tube.
25. Remove 384-well plate containing the dosed cells from the incubator.
26. Carefully remove the liquid from each well by gently tapping the plate several times upside down on a sterile surface. Ensure that all media is removed but do not disturb the cells.
27. Pour contents of 15 ml conical tube (containing CellTiter-Fluor™ reagent prepared in step #24) into a sterile media trough. Using a multi-channel pipette or liquid handler, gently add 25 µl of CellTiter-Fluor™ reagent into each well.
28. Place 384-well plate in the humidified 5% CO₂ /37°C incubator for 60 min.

The Following Steps Do Not Require Sterile Conditions

29. Remove 384-well plate from the incubator, briefly allow to cool to ambient temperature, and measure fluorescence of individual wells with a microplate reader in fluorescence mode using an excitation wavelength of 380-400 nm and an emission wavelength of 505 nm. Read plate at 1 msec/well.

30. Pour ONE-Glo™ Assay reagent (prepared in Step #23) into a media trough, and then add 25 µl of this reagent into each well of the plate.
31. Carefully agitate the plate to mix the reagents contained in the wells.
32. After 5 min, read the luminescence of individual wells using a luminometer set for a 5 sec pre-shake with a 1 sec/well read time. A very high gain (sensitivity) setting should be used.

DAY 3 P450 Metabolism and Receptor Activation (DPX2-384-002 kit)

THE FOLLOWING STEPS ARE PERFORMED IN A STERILE HOOD

33. Allow Tubes E, F, G, H, I, J & K to reach room temperature.
34. Mix 10 µl from Tube K (Luciferin-IPA) with 10 ml of Tube D (Dosing Medium) in a sterile 15 ml conical tube and pour in media reservoir.
35. Remove 384-well plate from the incubator after 48 h and place into the hood.
36. Carefully remove the liquid from each well by gently tapping the plate upside down several times on a sterile surface. *Ensure that all medium is removed but do not disturb cells.*
37. Add 25 µl from the media reservoir containing dosing medium with Luciferin-IPA reagent (prepared in step #34) to each well.
38. Return plate to incubator for 60 min at 37°C.
39. During incubation with Luciferin-IPA, prepare a replicate non-sterile white 384 well plate. In addition, pour the contents of Tube H (P450-Glo™ Buffer) into Tube I (Luciferin Detection Reagent). Mix by inversion. Pour into a media trough.
40. After the 60 min incubation, remove 384-well plate from the incubator. Carefully transfer 25 µl from each well of the original plate to the corresponding well of the white replicate non-sterile 384-well plate. After addition, remove non-sterile plate from hood.
41. Following transfer of medium containing the Luciferin-IPA reagent to replicate plate, transfer 10 ml from Tube E to a sterile 15 ml conical tube, and then add 5 µl from Tube J (CellTiter-Fluor™). Mix by inversion. Pour into media trough.
42. Using a multi-channel pipette or liquid handler, gently add 25 µl of CellTiter-Fluor™ reagent into each well of the original 384-well plate containing the cells (this is the plate that is still in the sterile hood). Return plate to the incubator for 60 min.

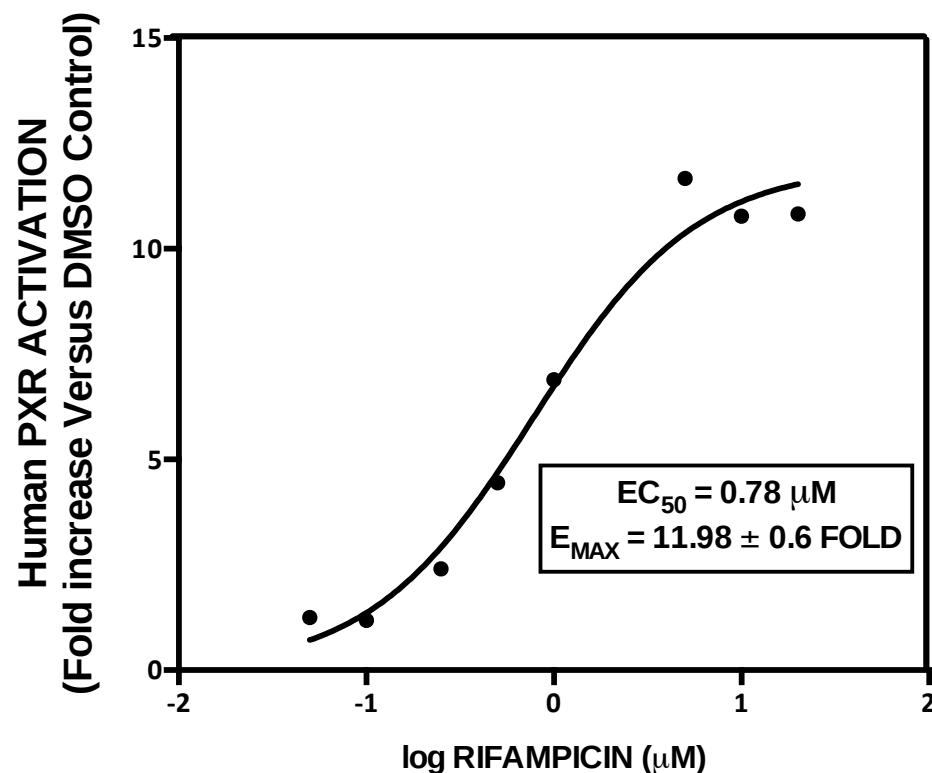
The Following Steps Do Not Require Sterile Conditions

43. Continuing with the replicate plate out side the hood, add 25 µl of P450-Glo™ Buffer/Luciferin Detection Reagent (prepared in step #39) to each well of the replicate plate, and incubate at room temperature for 20 min.
44. After the 20 min incubation with Luciferin Detection Reagent, measure the luminescence of the replicate 384-well plate with the luminometer set for a 1 sec/well read time. A fairly high gain setting should be used.

45. Add the contents of Tube F (ONE-Glo™ Assay Buffer) to Tube G (ONE-Glo™ Assay Substrate), cap, and mix by inversion.
46. Following the 60 min incubation with CellTiter-Fluor, remove the original 384-well plate from the incubator, allow to cool to room temperature, and then measure fluorescence of individual wells with a microplate reader in fluorescence mode using an excitation wavelength of 380-400 nm and a emission wavelength of 505 nm.
47. Pour ONE-Glo™ Assay reagent (prepared in Step #45) into a media trough, and add 25 µl of the reagent into each well of the plate.
48. Carefully agitate the plate to mix the reagents contained in the wells.
49. After 5 min, read the luminescence of the individual wells with the luminometer set for a 5 sec pre-shake with a 1 sec/well read time. A very high gain (sensitivity) setting should be used.

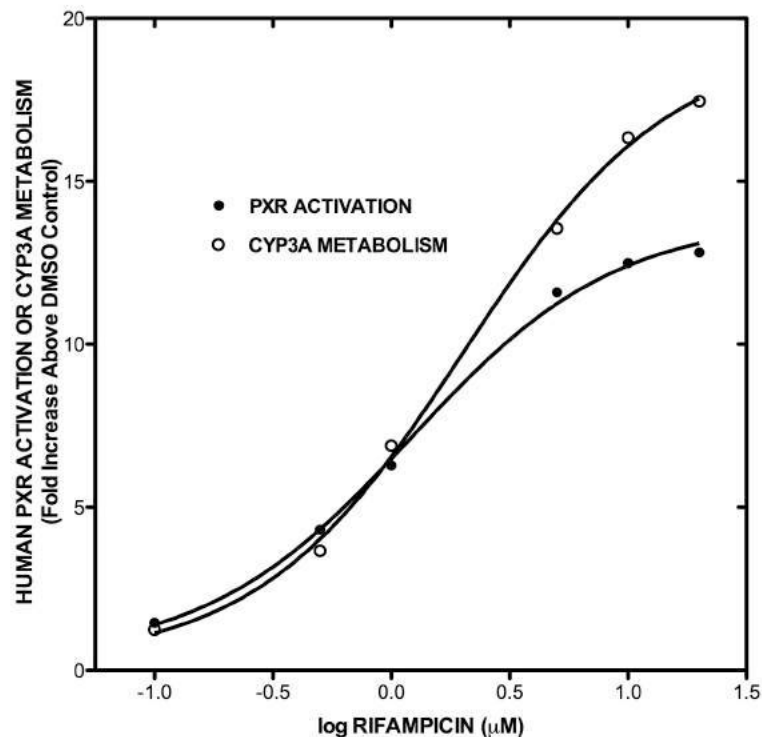
QUANTITATION OF PXR RECEPTOR ACTIVATION

1. Determine the average Relative Luminescence Units (RLU) of the three replicates for each test compound at each dosage. In addition, determine the average Relative Fluorescence Units (RFU) of the three replicates for each test compound at each dosage.
2. Determine the mean RLU and RFU for the DMSO vehicle control replicates.
3. Normalize the luciferase activity for cell viability by dividing the average RLU by the average RFU for each test compound at each dosage as well as for the vehicle control.
4. PXR receptor activation at the individual test compound doses is calculated by dividing the normalized luciferase activity (RLU/RFU) for the test compound doses by that of the normalized DMSO vehicle control. The final data is expressed as fold activation relative to the vehicle control. The use of 5 or more doses of test compound and positive control allows for the derivation of EC₅₀ and E_{MAX} values from nonlinear regression analysis of the log dose-response curves. Graphpad Prism (Graphpad Software, La Jolla, CA) and SigmaPlot (Systat Software, San Jose, CA) are among the software programs with built-in equations for deriving these PXR activation kinetic parameters.
5. Test compound values should be compared to those obtained for the positive (rifampicin) control. An example of a typical PXR activation dose-response curve generated with 8 concentrations of rifampicin is shown below.



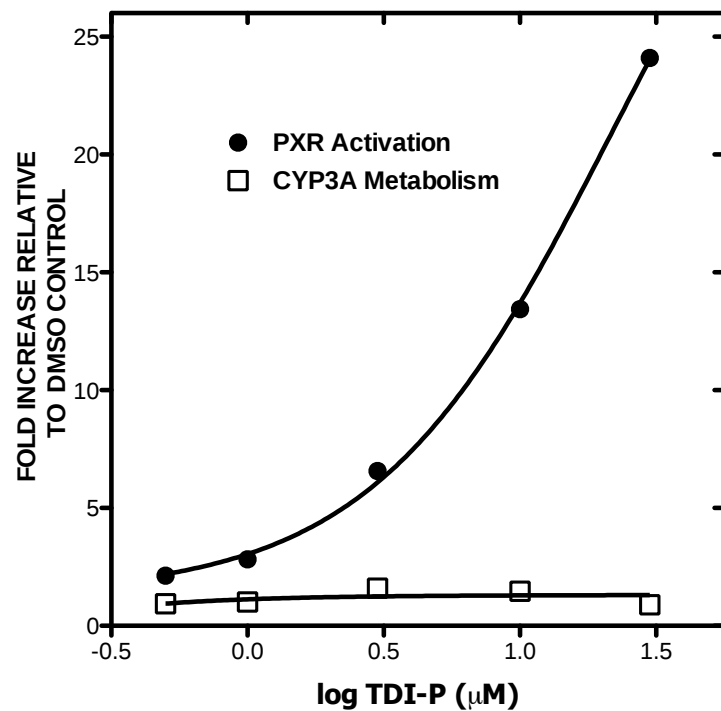
Quantitation of P450 Substrate Metabolism

1. Receptor activation by test compounds, positive controls, and negative (vehicle) controls are determined as described above.
2. To assess whether the test compounds also increased P450 metabolic activity, determine the average RLU of the three Luciferin-IPA replicates for each test compound at each dosage in the non-sterile replicate white 384-well plates. In addition, determine the average RLU of the three replicates for the DMSO (vehicle) controls.
3. Normalize the P450-Glo™ activity for cell viability by dividing the average RLU by the average viability (RFU) determined in the original plate for each test compound at each dosage as well as for the vehicle control.
4. Fold induction by the individual doses of test compound is calculated by dividing the normalized P450-Glo™ luciferase activity (RLU/RFU) for the test compound doses by that of the normalized DMSO vehicle control. The results should be expressed as fold increase (induction) above DMSO-treated cells.



A typical dose-response curve generated in DPX2™ cells treated with 6 concentrations of rifampicin. Both PXR activation and P450 3A metabolism of Luciferin-IPA were monitored in a multiplex fashion. As expected, both metabolic activity and PXR activation were enhanced by rifampicin.

NOTE: It is not uncommon to obtain extensive PXR activation but negligible induction of P450 substrate metabolism. This happens in cases where the PXR activating compound inhibits the P450 enzyme that metabolizes the P450 Glo™ substrate. The capacity to multiplex Puracyp's Human PXR Activation Assay System allows one to identify such molecules (see p. 12) as well those that exhibit cellular toxicity.



Dose-response curve generated in DPX2[™] cells treated with the metabolic inhibitor TDI-P. Both PXR activation and CYP3A metabolism of Luciferin-IPA were monitored in a multiplex fashion. The lack of an increase in Luciferin-IPA metabolism with increasing doses of TDI-P suggests that this compound is indeed a metabolic or time-dependent inhibitor of CYP3A.

TROUBLESHOOTING GUIDE

SYMPTOM	POSSIBLE CAUSE	RESOLUTION
High Well-to-Well Variation	Cells Not Evenly Dispersed Among Wells	Mix cells thoroughly with medium before seeding. To facilitate even plating, ensure the cells in Tube A are pipetted up/down at least twice prior to transfer to culture medium. Upon transfer, mix cells with culture medium by pipetting up/down 10 times.
Weak Activation with Potent Positive Control	Inadequate cell number	Ensure that there are 6 million viable cells per vial. Each well of a 384 well plate should receive 15,000 cells/well. To ensure that the cells arrived in good condition following shipping, it is recommended that the cells be counted and viability checked prior to seeding in the 384 well plate.
	Luminometer Settings	The luminometer used should be one that can be set to read 1 second/well at a very high sensitivity or gain.

Weak Activation with Test Compounds	Poor Test Compound Solubility	Test compounds dissolved in DMSO and other solvents precipitate out of solution when added to culture medium. This often occurs at higher concentrations. Weak activation stemming from poor solubility would be indicated by: a) a peak activation response occurring midway in the dose-response curve, followed by decreasing responses at higher concentrations and; b) no change in cell viability throughout the entire concentration curve.
	Effects on Cell Viability	Test compound cytotoxicity can result in weak PXR activation. Cell viability should be assessed using CellTiter-Fluor™ reagent, and the fold PXR activation normalized to the viable cell number for a given test compound. A decline in receptor activation and cell viability over the concentration range examined would indicate that the test compound is toxic to the DPX2™ cells.
False Positives	Borderline Activation by Non-PXR Ligands	The low levels of PXR activation observed with certain compounds may not be indicative of “false positives”. Cut-off values, commonly expressed as % of RIF, should be established to differentiate negative, weak, moderate and potent PXR activators. For example, compounds giving activation < 15% of that observed with 10 µM RIF would be considered negative. Agents giving 15 - 40% of the response observed with 10 µM RIF would be considered weak activators while those eliciting 41 - 69% of the RIF response would be moderate activators. Only those compounds giving ≥ 70% of the response noted with 10 µM RIF would be considered strong or potent PXR activators.

Potent PXR Activation But Weak Induction of CYP3A Metabolism	Test Compound is an Irreversible (Time-Dependent) CYP3A Inhibitor	Certain agents elicit potent activation of PXR together with marked induction of CYP3A enzyme levels but fail to give a corresponding increase in CYP3A-dependent metabolism. This well-documented phenomenon stems from the irreversible, time-dependent inhibition of CYP3A P450s by such compounds during their metabolism by the same enzyme(s). An example is provided on p.12 of this User's Manual.
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For any additional questions and/or problems with Puracyp's *Human PXR Activation Assay System*, please contact us by phone at 760-929-9508 or by email at support@puracyp.com.

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DPX2™ cells are covered by US PATENT NO. 6,984,489 owned by Xenopharm Inc. and licensed by Puracyp.

ONE-Glo™ and CellTiter-Fluor™ are subject to patents pending by Promega Corporation.

P450-Glo™ CYP3A4 Assay with Luciferin-IPA is covered by U.S. Pat. Nos. 6,602,677 and 7,724,584, European Pat. No. 1131441 and other patents pending owned by Promega Corporation.

The method of recombinant expression of Coleoptera luciferase is covered by U.S. Pat. Nos. 5,583,024, 5,674,713 and 5,700,673.

Additional information on ONE-Glo™, CellTiter-Fluor™ and the P450-Glo™ CYP3A4 Assay with Luciferin-IPA can be found at www.promega.com.

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COMPOUNDS THAT ARE TOXIC OR INHIBIT CELL CYCLE EVENTS MAY NOT EXHIBIT INDUCTION IN PURACYP'S CELL LINES BUT RECEPTOR ACTIVATION MAY BE OBSERVED IF A VIABILITY ASSAY IS PERFORMED ON COMPOUNDS THAT ARE TOXIC AND RESULTS NORMALIZED TO THAT ASSAY.

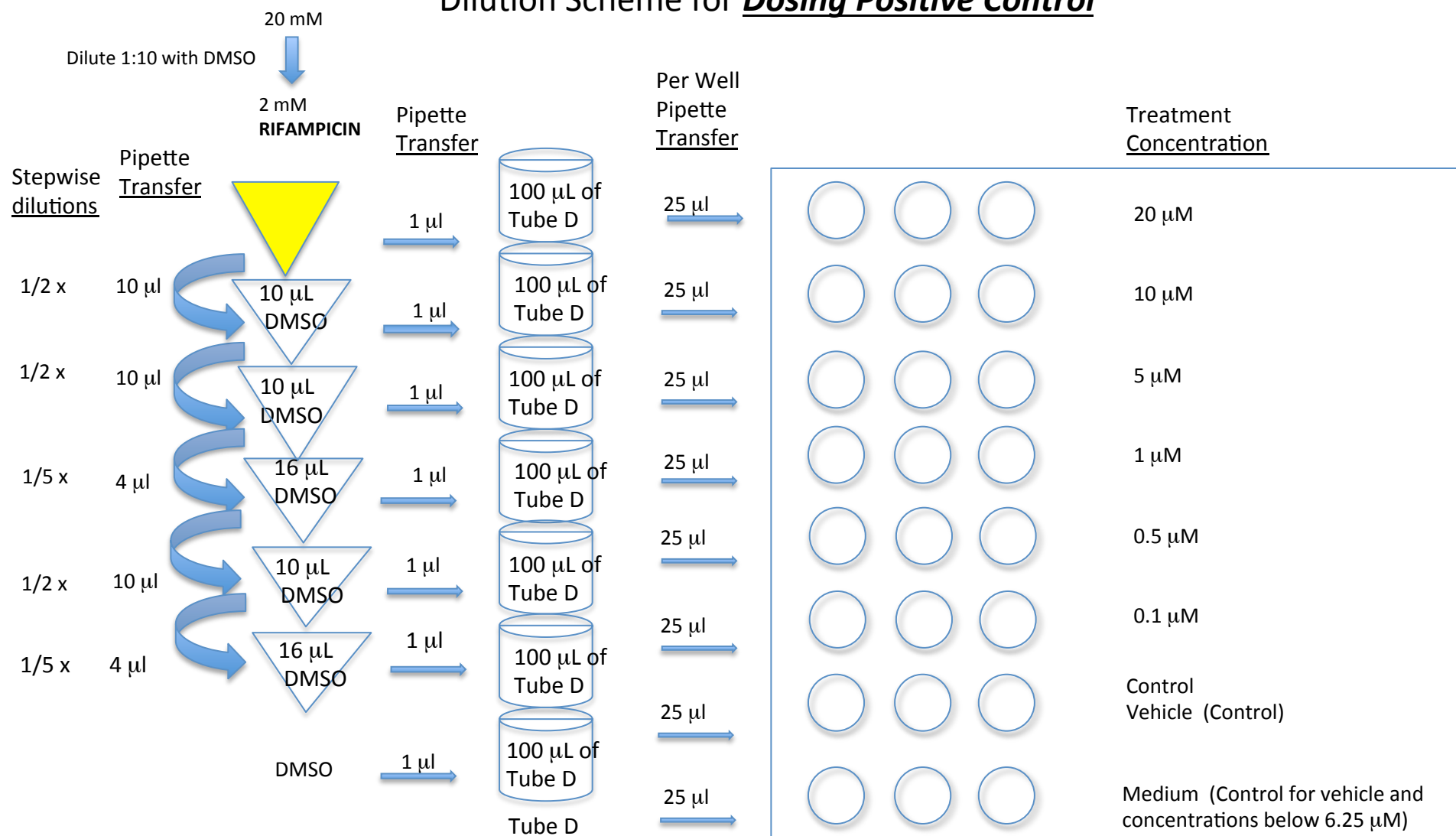
IF THE ABOVE TERMS ARE NOT ACCEPTABLE, PROMPTLY CONTACT PURACYP. FAILURE TO PROMPTLY CONTACT PURACYP IN WRITING WITHIN TWO DAYS OF DELIVERY CONSTITUTES ACCEPTANCE OF THE ABOVE TERMS. DEVIATIONS FROM THE ABOVE PROTOCOLS WILL VOID WARRANTIES.

VI. REFERENCES

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APPENDIX 1 FOR DPX2-384-001 AND DPX2-384-002

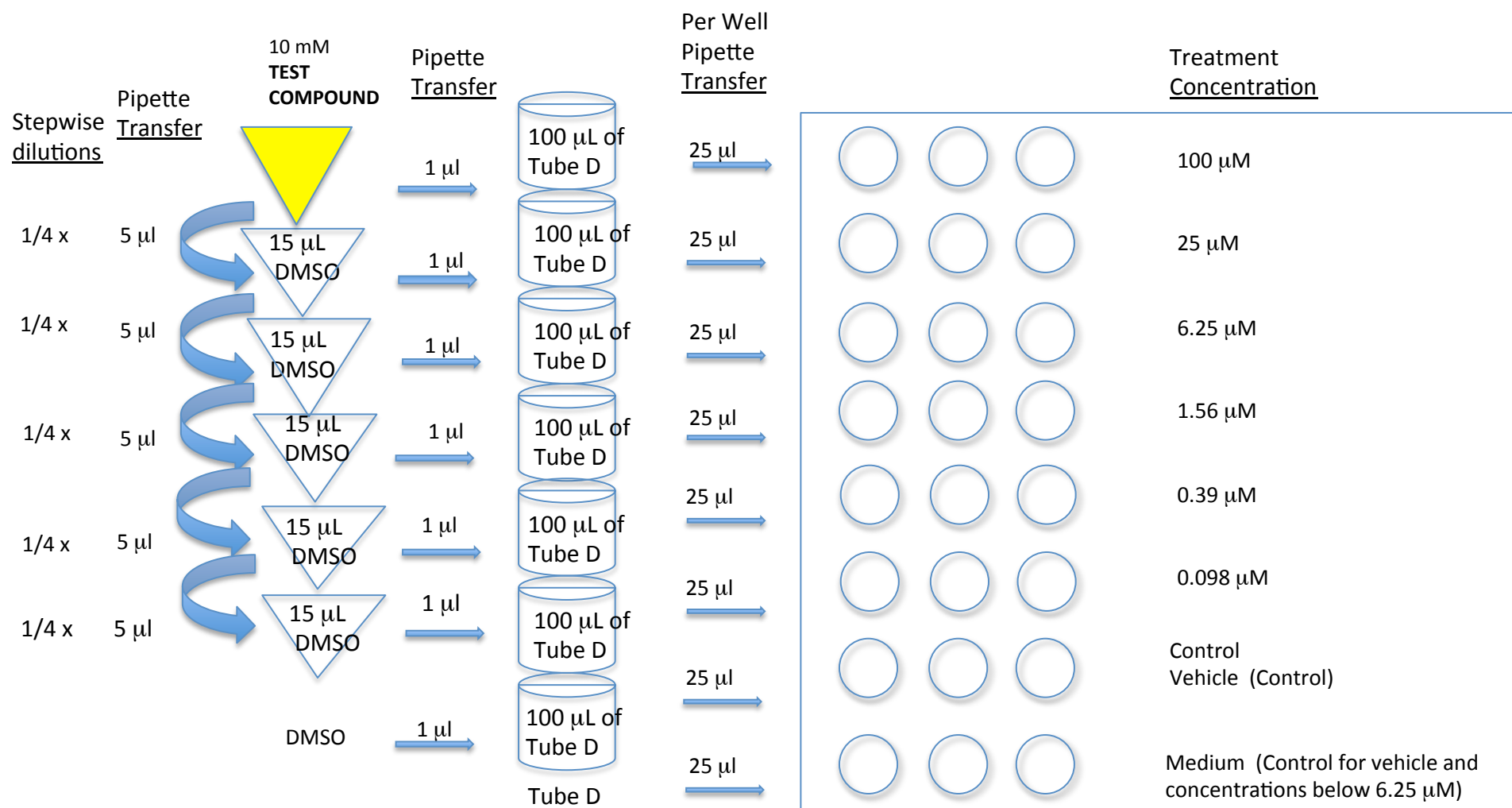
Dilution Scheme for *Dosing Positive Control*



All test compound dilutions are made with neat DMSO, after which the individual solutions are added to dosing media at a ratio of 1:100.

APPENDIX 2 FOR DPX2-384-001 AND DPX2-384-002

Dilution Scheme for Dosing Test Compounds



All test compound dilutions are made with neat DMSO, after which the individual solutions are added to dosing media at a ratio of 1:100.

APPENDIX 3: QUICK REFERENCE CARD

HUMAN PXR NUCLEAR RECEPTOR ACTIVATION ASSAY WITH ASSESSMENT OF CELL VIABILITY IN A 384-WELL FORMAT (DPX2-384-001)

USE STERILE TECHNIQUE FOR STEPS 1-17

KIT RECEIPT DAY

1. Thaw Tube C by warming in a 37°C water bath, and transfer 10 ml into a media trough.
2. Place Tube A containing DPX2 cells in a 37°C water bath until cell suspension has thawed.
3. Pipette cells into the media trough containing 10 ml of Reagent C, and mix by pipetting up-and-down 10 times. Count cells and determine viability.
4. Transfer 25 µl of cell mixture into plate wells, keeping cells suspended while pipetting.
5. Place seeded plate into a humidified CO₂ incubator at 37°C overnight.

DAY 1

6. Thaw Tube D in a 37°C water bath.
7. Allow Tube B to thaw at room temperature.
8. Prepare series of Test Compound and Positive Control dilutions in DMSO (see appendices 1 & 2) Followed by 1:100 dilutions into Dosing Medium (Tube D).
9. Remove plate from the incubator, and remove media from each well by gently tapping the plate upside down on sterile surface. Do not disturb the cells.
10. Transfer 25 µl of each test compound concentration into pre-marked wells. Do the same with positive controls and vehicle controls.
11. Return plate to the incubator for a 48 h exposure to the treatments.

DAY 3

12. Thaw Tube J containing CellTiter-Fluor™.
13. Thaw Tube E, transfer 10 ml to a 15 ml sterile centrifuge tube, and add 5 µl of CellTiter-Fluor™ (Tube J). Mix by inversion and pour into a media trough.
14. Thaw Tube F containing ONE-Glo™ Luciferase Assay Buffer, and add to Tube G containing ONE-Glo™ Assay Substrate; mix vial several times by inversion. Pour into media trough.
15. Remove plate from the incubator, and discard all dosing medium containing test compounds, positive controls, and negative (vehicle) controls by gently tapping the plate upside down on a sterile surface.
16. Add 25 µl of Tube E containing the CellTiter-Fluor™ reagent (prepared in Step 13) to each well.
17. Return plate to the incubator for 60 min.
18. After incubation, measure fluorescence of individual wells with a multi-mode microplate reader^A set for fluorescence using an excitation wavelength of 380-400 nm and an emission wavelength of 505 nm.
19. Remove plate from instrument, and add 25 µl of ONE-Glo™ Assay reagent to each well. Agitate plate to mix, and incubate for 5 min at room temperature.
20. Measure luminescence of individual wells with the multi-mode reader set for a 5 sec pre-shake and a 1 sec/well read. A high instrument gain (sensitivity) setting should be used.
21. Receptor activation is calculated according to the method given in the *Complete Protocol*.

CellTiterFluor™ and ONE-Glo™ are trademarks of the Promega Corporation.

^AA stand-alone fluorimeter and luminometer can also be used

APPENDIX 4: QUICK REFERENCE CARD
HUMAN PXR NUCLEAR RECEPTOR ACTIVATION ASSAY WITH CELL VIABILITY AND CYP3A
SUBSTRATE METABOLISM IN A 384-WELL FORMAT (DPX2-384-002)

USE STERILE TECHNIQUE FOR STEPS 1-20

KIT RECEIPT DAY

1. Thaw Tube C by warming in a 37°C water bath, and transfer 10 ml into a media trough.
2. Place Tube A containing DPX2 cells in a 37°C water bath until cell suspension has thawed.
3. Pipette cells into the media trough containing 10 ml of Reagent C, and mix by pipetting up-and-down 10 times. Count cells and determine viability.
4. Transfer 25 µl of cell mixture into plate wells, keeping cells suspended while pipetting.
5. Place seeded plate into a humidified CO₂ incubator at 37°C overnight.

DAY 1

6. Thaw Tube D in a 37°C water bath.
7. Allow Tube B to thaw at room temperature.
8. Prepare series of Test Compound and Positive Control dilutions in DMSO followed by 1:100 dilutions into dosing medium (Tube D) (see appendices 1 & 2). Remaining contents of Tube D should be stored at 4°C. Ten ml of Tube D will be needed for the metabolism assay (see step # 14 below).
9. Remove plate from the incubator, and discard the media from each well by gently tapping the plate upside down on a sterile surface. Do not disturb the cells.
10. Transfer 25 µl of each test compound concentration into pre-marked wells. Repeat with positive controls and vehicle controls.
11. Return plate to incubator for 48 h^A

DAY 3

12. Thaw Tubes J and K containing CellTiter-Fluor™ and Luciferin-IPA, respectively.
13. Allow Tubes D, E, F, G, H, and I to reach room temperature.
14. Add 10 µl of Tube K to 10 ml of Tube D. Mix by inversion. Pour into media trough.
15. Remove 384-well plate from incubator and carefully remove media from wells by gently tapping the plate upside down on a sterile surface.
16. Add 25 µl of the dosing medium containing Luciferin-IPA reagent (prepared in step #14) to each well. Return plate to incubator for 60 min.
17. During the incubation, pour Tube H (P450-Glo buffer) into Tube I (Luciferin Detection Reagent) and mix by inversion. Pour into a media trough.
18. After 60 min, remove 384-well plate from the incubator, and transfer 25 µl from each well to a corresponding well of a non-sterile white 384-well plate that replicates the format of the original plate.
19. After transfer of Luciferin-IPA reagent to replicate plate, transfer 10 ml of Tube E to a 15 ml sterile conical tube, add 5 µl of CellTiter-Fluor™ (Tube J), and mix by inversion. Pour into media trough.

20. Using a multi-channel pipette or liquid handler, gently add 25 μ l of CellTiter-Fluor™ reagent into each well of the original 384-well plate containing the cells. Return plate to the incubator for 60 min.
21. Add 25 μ l of Luciferin Detection Reagent/P450-Glo Buffer (prepared in step #17) to each well of the replicate micro-titer plate, agitate the plate, and incubate at room temperature for 20 min.
22. After 20 min incubation with Luciferin Detection Reagent, determine luminescence of the wells of the non-sterile white 384-well plate with the luminometer set for a 1 sec/well read time. A relatively high gain setting should be used.
23. Add Tube F containing ONE-Glo™ Luciferase Assay Buffer to Tube G containing ONE-Glo™ Assay Substrate; mix by inversion. Pour into media trough.
24. After 60 min incubation at 37°C (see step #20), remove original 384-well plate from incubator and measure fluorescence of individual wells with multi-mode microplate reader^B set to the fluorescence mode using an excitation wavelength of 380-400 nm and an emission wavelength of 505 nm.
25. Remove plate from instrument, and add 25 μ l of ONE-Glo™ Assay reagent (prepared in step # 23) to each well. Agitate plate to mix, and incubate for 5 min at room temperature.
26. Measure luminescence of individual wells with the multi-mode reader set for a 5 sec pre-shake and a 1 sec/well read. A high instrument gain (sensitivity) setting should be used.
27. Receptor activation is calculated according to the method given in the *Complete Protocol*.

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^APuracyp's Human PXR Nuclear Receptor Activation Assay is designed to give maximum induction upon treatment of DPX2 cells for 48 h at 37°C with a single dose of RIF. However, compounds other than RIF may be less stable in culture at 37°C, and thus may not function as PXR ligands for the entire treatment period. In those instances, the user may want to perform two 24 h treatments, which will require that Day 1 Steps #8-11 are repeated on Day 2.

^BA stand-alone fluorimeter and luminometer can also be used.